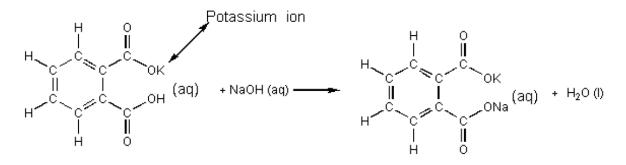
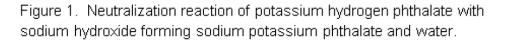
## **STANDARDIZATION OF NaOH**

<u>Purpose:</u> You will precisely measure the concentration of the NaOH solution you prepared last lab by carrying out a series of acid/base neutralizations.

<u>Background:</u> A **neutralization** is when an acid and a base react together to form a salt. You will react a strong base, sodium hydroxide, with a weak acid, potassium hydrogen phthalate (Figure 1). Because potassium hydrogen phthalate is awkward to say/write, we abbreviate the name KHP. This is confusing because it looks like a chemical formula, but it's not. There is no phosphorus (P), and remember that potassium's symbol is K, not P. The products formed are KNaP (potassium sodium phthalate) and water.





As described in our textbook, when you want to know exactly how much of a substance you have, you can carry out a titration. Using an exact amount of one reactant (the **titrant**) you can figure out how much of the other reactant (the **analyte**) must be present. There has to be some obvious sign when the mole ratio of titrant and analyte match the balanced equation ratio. This is the **equivalence point**. We measure the equivalence point by adding an **indicator** molecule that changes color. The human eye is much better at telling the difference between "clear" and "colored" than it is at distinguishing between two different shades, so the endpoint (when you should stop) is when the solution is the faintest shade of color that you can see (Figure 2). At this point, the balanced equation tells you the ratio of reactants; neither one is in excess. Titrations are very useful in medicine (determining how much medication someone needs), food processing (figuring out how much fat is in a product), and industry (measuring how much heavy metal is in a waste stream).

For the NaOH solution you made last lab, only the approximate concentration is known. You did not use exact volumes (the bottle used is not an exact volumetric piece of glassware), and the deionized water you used contains some impurities (like carbon dioxide). You will measure the exact concentration of NaOH using pure KHP as the titrant. Next lab, you will turn around and use the exact concentration of this standardized solution. Look at one of these tutorial videos for titration tips: <u>https://www.youtube.com/watch?v=sFpFCPTDv2w</u> or https://www.youtube.com/watch?v=9DkB82xLvNE



Figure 2: Colors observed during a titration. Don't go too far!

Start Endpoint Too Far

Procedure: This is the part you have to write in your notebook. You will work alone for this lab.

- 1. Obtain from the desiccator one of the vials of KHP. The molecular weight of KHP is 204.23 g/mol.
- 2. Obtain a buret and the bottle of NaOH you made last week from the cabinet. Make sure your buret is clean and the stopcock is tightened. Rinse the buret three times with 5 mL portions of your NaOH solution.
- 3. Fill the buret, and adjust the volume between 0 and 1 mL. Remove any bubbles; don't forget about the tip of the buret. Read and record the initial volume to two decimal places. Make sure your eye is lined up to avoid parallax errors.
- 4. Use the analytical balance to weigh at least 1.0 g but not more than 1.1 g of KHP. Record all 4 decimal places. Transfer the weighed KHP to a 250 mL Erlenmeyer flask, and use deionized water from your squirt bottle to rinse any remaining solid in the weigh boat into the flask.
- 5. Dissolve the KHP in 75 mL of deionized water. Don't worry if not all of it dissolves, just write it down as an observation, and make sure it all dissolves before the endpoint. Add 3 drops of phenolphthalein indicator to the flask.
- 6. Dispense NaOH from the buret into the flask and swirl. When the solution turns faint pink and the color lasts 10 seconds, you have reached the endpoint. It should take 20 40 mL of NaOH. Write the final volume down to two decimal places. Expert titrators can split drops to get exactly the endpoint. If it turns dark pink (fuscia) you have overshot the endpoint. After the titration, the solution may be rinsed down the drain.
- Repeat the titration 3 times. Keep the mass of KHP used as constant as possible. Calculate the concentration of your NaOH solution and the <u>average deviation</u> (as in Lab 1). If the average deviation is less than 0.0002 M, you can stop. If not, do another trial or two (no more than 5 total).
- 8. Before the buret is stored, rinse it thoroughly with water, and loosen the stopcock. Put the KHP vial in the box **not** in the desiccators. Put the bottle of NaOH back in the cabinet for the next lab.

## **Example Data Table:**

	Trial 1	Trial 2	Trial 3	Trial 4	Trial 5
Mass KHP					
Initial buret vol					
Final buret vol					

After completing the procedure but before leaving lab, write in your notebook a brief statement (two to three sentences) on the quality and reasonableness of the data you collected. Note what you might do differently if you performed the lab again.

## **Tips:**

- Do not throw the data out if you overshoot. One extra drop can cause a huge change in color but have a pretty small effect on concentration. Just record it as an observation.
- Use the deionized water squirt bottle to rinse down the sides of your flask if anything splashes.
- Do the first run quickly to get an idea of where the endpoint will be. Then use similar masses of KHP for the subsequent runs, so you'll know exactly when to slow down.
- If the volume required to titrate 1 g of KHP is not in the range of 20-50 mL of NaOH, your solution was made incorrectly (you probably used KOH or measured the volume incorrectly). Talk to your instructor.
- Watch significant figures. Rounding errors can affect your precision.
- If you've added more base than should be required and there's still no hint of pink, make sure you didn't forget the phenolphthalein indicator.
- If you're not sure if something is pink, try comparing it to a white sheet of paper near natural light.